

Viktor Kettmann,^{a*} Jan Lokaj,^b
Christoph Kratky,^c Stefan
Marchalin^b and Jana Sikoraiova^b^aFaculty of Pharmacy, Comenius University, Odbojarov 10, Bratislava 83232, Slovak Republic, ^bFaculty of Chemical Technology, Slovak Technical University, Radlinskeho 9, Bratislava 81237, Slovak Republic, and ^cInstitut für Physikalische Chemie, Karl-Franzens-Universität Graz, Heinrichstrasse 28, Graz 8010, AustriaCorrespondence e-mail:
kettmann@fpharm.uniba.sk

Key indicators

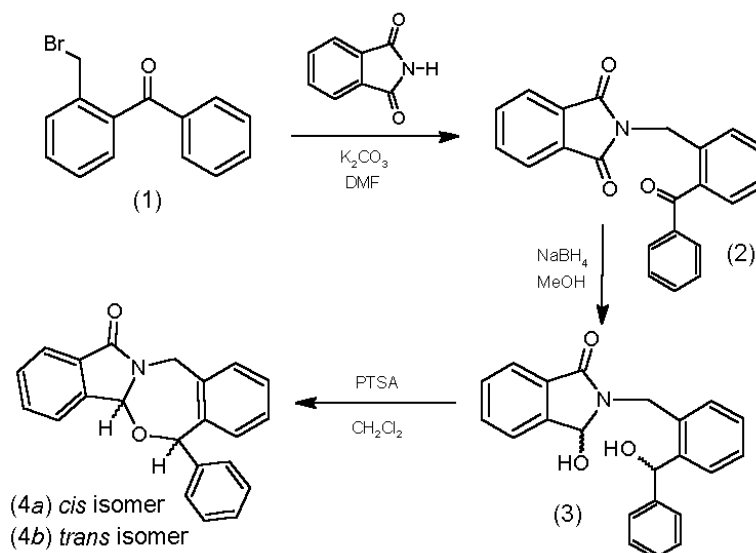
Single-crystal X-ray study
T = 293 K
Mean $\sigma(\text{C}-\text{C}) = 0.004 \text{ \AA}$
R factor = 0.074
wR factor = 0.194
Data-to-parameter ratio = 12.9For details of how these key indicators were automatically derived from the article, see <http://journals.iucr.org/e>.*trans*-5,6a-Dihydro-5-phenylisoindolo[1,2-*b*]-
benz[1,3]oxazepin-11-one

The title compound, $\text{C}_{22}\text{H}_{17}\text{NO}_2$, is composed of an oxoisoindoline moiety fused to a phenyl-substituted benzoxazepine ring and was designed and synthesized as a potential anxiolytic drug. The isoindolinone moiety is essentially planar and the central oxazepine ring adopts a twist-boat conformation with the phenyl group equatorial. In the two independent molecules, the benzene ring of the benzoxazepine fragment makes an angle of $74.4(1)$ or $86.1(1)^\circ$ with the plane of the isoindoline ring.

Received 22 May 2001
Accepted 14 June 2001
Online 22 June 2001

Comment

This work is part of our continuing study aimed at designing modulators of hormonal/neurotransmitter systems as potential drugs to treat neuronal and cardiovascular disorders. Based on the recent pharmacophore/receptor model of the benzodiazepine (BDZ) receptor subtype located in the central nervous system (Huang *et al.*, 2000), we designed the compound (4), as potential anxiolytic agent. Synthesis of the molecule (4) was achieved by a sequential reaction and led to a 5:1 diastereomeric (racemic) mixture of *cis*-(4a) and *trans*-(4b) isomers. In order to establish the detailed stereochemistry of the two diastereomers, *viz.* spatial relationship between the putative pharmacophoric elements (phenyl rings and the two O atoms) which is indispensable for future molecular-modelling studies, the crystal structure determination of (4a) and (4b) has been undertaken. We report here on the structure of the *trans*-(4b) isomer.



In *trans*-(4*b*), two independent molecules (*A* and *B*) are identical to within 4σ as far as bond distances and angles are concerned. Thus, only one molecule (*A*) along with the atom-numbering scheme is shown in Fig. 1. As expected, the isoindolinone ring is essentially planar. The N1—C2 bond is much shorter than the N1—C9 and N1—C18 bonds (Table 1); such N—C bond lengths resemble those typically found in cyclic amino acids (Benedetti *et al.*, 1983), indicating that the lone-pair electrons on N1 are involved in conjugation with the adjacent carbonyl group. A similar pattern of bond distances and angles within the isoindolinone moiety has been found in the *cis* isomer (Lokaj *et al.*, 2001) as well as other compounds incorporating this molecular fragment (Barrett *et al.*, 1995; McNab *et al.*, 1997; Khan *et al.*, 1998).

As mentioned above, the main purpose of this structure determination was to establish the relative three-dimensional disposition of the phenyl rings and the two O atoms which are assumed to constitute the interaction pharmacophore responsible for binding of the compound to the CNS-subtype of the BDZ receptor. Obviously, the disposition of these structural elements depends primarily on the conformation of the seven-membered oxazepine ring which is the most flexible part of the molecule. A comparison of the endocyclic torsion angles for the oxazepine ring (Table 1) reveals that the ring adopts a twist-boat conformation with an approximate twofold axis passing through C9 and the midpoint of the C12—C17 bond.

The puckering parameters according to Cremer & Pople (1975) are $q_2 = 0.817(4) \text{ \AA}$, $\varphi_2 = 175.7(3)^\circ$ and $q_3 = 0.370(4) \text{ \AA}$, $\varphi_3 = 106.8(5)^\circ$ for the sequence N1A/C9A/O10A/C11A/C12A/C17A/C18A. The corresponding parameters in molecule *B* are $q_2 = 0.814(4) \text{ \AA}$, $\varphi_2 = 169.2(3)^\circ$ and $q_3 = 0.378(4) \text{ \AA}$, $\varphi_3 = 108.8(6)^\circ$. The deviation from ideal C_2 symmetry described by the asymmetry parameter $\Delta C_2(C9)$ is 0.062(1) (molecule *A*) and 0.026(1) (molecule *B*) (Nardelli, 1983). Although the puckering mode of the oxazepine ring in molecules *A* and *B* is the same, the endocyclic torsion angles in the two molecules differ by up to 23σ . Another difference between molecules *A* and *B* concerns the orientation of the phenyl group (at C11) as shown by the torsion angle O10—C11—C19—C24 which is $46.3(3)^\circ$ in *A* and $64.0(3)^\circ$ in *B*. This points to the flexibility of the oxazepine ring and the shallow shape of the potential well corresponding to rotation of the phenyl group about the exocyclic C11—C19 bond. In both molecules, of course, the phenyl substituent occupies a pseudo-equatorial position. The equatorial arrangement of the phenyl group has also been observed for the *cis* isomer (Lokaj *et al.*, 2001) but in the latter compound the oxazepine ring exists in a distorted C9-chair conformation. This is in line with the known fact that the equatorial orientation of bulky substituents attached to a saturated (or partially unsaturated) seven-membered ring is more important than the actual conformation of the ring, obviously due to low barriers along the pseudorotation pathway. Due to the relatively severe puckering of the central seven-membered ring, the molecules as a whole are non-planar: the two planar 'ends' (*viz.* the isoindoline and the benzene ring of the benzoxazepine

moiety) are inclined at an angle of $74.4(1)$ and $86.1(1)^\circ$ in molecules *A* and *B*, respectively. A similar molecular shape has been found for the *cis*-isomer [bent angle $67.7(1)^\circ$]. This implies an equivalent spatial relationship between the pharmacophoric elements in the two diastereomers and hence, based on the crystal structure data, a similar pharmacological behaviour for the two isomers is predicted.

Experimental

The diastereomers (4*a*) and (4*b*) were synthesized by a three-step reaction. As the first step, to bromomethylbenzophenone (1), prepared freshly from 2-methylbenzophenone (1.96 g, 0.01 mol) and *N*-bromosuccinimide (1.76 g, 0.01 mol), was added phthalimide (1.5 g, 0.01 mol), potassium carbonate (1.1 g, 8 mmol) and *N,N*-dimethylformamide (25 ml). The mixture was stirred overnight, diluted with water, extracted with diethyl ether (3×20 ml) and dried (magnesium sulfate). The solvent was evaporated under reduced pressure and the solid recrystallized from ethanol to give 2-(*N*-phthalimidomethyl)benzophenone, (2) (77% yield, m.p. 388 K). In the second step, to a solution of (2) (0.5 g, 15 mmol) in dry methanol (20 ml) at 273–283 K was added sodium borohydride (0.69 g, 30 mmol) by portions. The mixture was stirred for 2 h and monitored by TLC (dichloromethane/acetone 5:1). After 2 h, the starting material disappeared and the excess of sodium borohydride was decomposed by addition of cold water (10 ml) and 10% hydrochloric acid to neutral pH. The precipitate was separated by filtration, washed with water, dried, concentrated under reduced pressure and recrystallized from ethanol to afford a 5:1 ratio of diastereomers (3) (79% yield). Finally, compound (4) was prepared from the diols (3) (0.5 g, 1.5 mmol) were stirred in dry dichloromethane (20 ml) with a catalytic amount of *p*-toluenesulfonic acid for 30 min at room temperature. The solution was washed with saturated sodium hydrogen carbonate, with water, then dried and concentrated under reduced pressure. Separation of the product by flash chromatography and recrystallization from ethanol gave the corresponding 5:1 ratio of oxazepines (4*a*) and (4*b*) (70% yield); m.p. (4*a*) 496 K and (4*b*) 482 K. The isomers were initially characterized by IR, ^1H and ^{13}C NMR spectral analyses.

Crystal data

$\text{C}_{22}\text{H}_{17}\text{NO}_2$	$D_m = 1.30(1) \text{ Mg m}^{-3}$
$M_r = 327.37$	D_m measured by flotation in bromoform/cyclohexane
Triclinic, $P\bar{1}$	Mo $K\alpha$ radiation
$a = 10.799(3) \text{ \AA}$	Cell parameters from 25 reflections
$b = 12.113(4) \text{ \AA}$	$\theta = 7\text{--}18^\circ$
$c = 13.775(4) \text{ \AA}$	$\mu = 0.08 \text{ mm}^{-1}$
$\alpha = 104.47(4)^\circ$	$T = 293(2) \text{ K}$
$\beta = 99.40(3)^\circ$	Prism, colourless
$\gamma = 101.26(5)^\circ$	$0.40 \times 0.30 \times 0.25 \text{ mm}$
$V = 1667.9(9) \text{ \AA}^3$	
$Z = 4$	
$D_x = 1.304 \text{ Mg m}^{-3}$	

Data collection

Siemens <i>P4</i> diffractometer	$h = -1 \rightarrow 12$
$\omega/2\theta$ scans	$k = -13 \rightarrow 13$
6826 measured reflections	$l = -16 \rightarrow 16$
5812 independent reflections	3 standard reflections
3954 reflections with $I > 2\sigma(I)$	every 97 reflections
$R_{\text{int}} = 0.050$	intensity decay: 2%
$\theta_{\text{max}} = 25.0^\circ$	

Refinement

Refinement on F^2
 $R[F^2 > 2\sigma(F^2)] = 0.074$
 $wR(F^2) = 0.194$
 $S = 1.04$
 5812 reflections
 451 parameters

H-atom parameters constrained
 $w = 1/[\sigma^2(F_o^2) + (0.1139P)^2]$
 where $P = (F_o^2 + 2F_c^2)/3$
 $(\Delta/\sigma)_{\max} = 0.001$
 $\Delta\rho_{\max} = 0.35 \text{ e } \text{\AA}^{-3}$
 $\Delta\rho_{\min} = -0.52 \text{ e } \text{\AA}^{-3}$

Table 1
 Selected geometric parameters (\AA , $^\circ$).

N1A—C2A	1.368 (4)	N1B—C2B	1.370 (4)
N1A—C9A	1.447 (3)	N1B—C9B	1.449 (3)
N1A—C18A	1.449 (3)	N1B—C18B	1.456 (3)
C2A—O2A	1.227 (3)	C2B—O2B	1.224 (3)
C2A—C3A	1.490 (4)	C2B—C3B	1.486 (4)
C3A—C8A	1.373 (4)	C3B—C8B	1.382 (4)
C8A—C9A	1.510 (4)	C8B—C9B	1.506 (4)
C9A—O10A	1.421 (3)	C9B—O10B	1.427 (3)
O10A—C11A	1.455 (3)	O10B—C11B	1.454 (3)
C11A—C19A	1.510 (3)	C11B—C19B	1.512 (3)
C11A—C12A	1.537 (3)	C11B—C12B	1.533 (3)
C12A—C17A	1.403 (3)	C12B—C17B	1.407 (3)
C17A—C18A	1.520 (4)	C17B—C18B	1.522 (4)
C2A—N1A—C9A	114.1 (2)	C2B—N1B—C9B	113.4 (2)
C2A—N1A—C18A	125.8 (2)	C2B—N1B—C18B	125.0 (2)
C9A—N1A—C18A	119.9 (2)	C9B—N1B—C18B	119.2 (2)
O2A—C2A—N1A	125.9 (3)	O2B—C2B—N1B	125.9 (3)
O2A—C2A—C3A	128.7 (3)	O2B—C2B—C3B	128.0 (3)
N1A—C2A—C3A	105.5 (2)	N1B—C2B—C3B	106.1 (2)
O10A—C9A—N1A	111.18 (19)	O10B—C9B—N1B	110.8 (2)
O10A—C9A—C8A	116.6 (2)	O10B—C9B—C8B	115.0 (2)
N1A—C9A—C8A	101.9 (2)	N1B—C9B—C8B	102.3 (2)
C9A—O10A—C11A	112.93 (18)	C9B—O10B—C11B	112.97 (18)
C18A—N1A—C9A—O10A	46.6 (3)	C18B—N1B—C9B—O10B	43.7 (3)
N1A—C9A—O10A—C11A	47.9 (3)	N1B—C9B—O10B—C11B	51.1 (3)
C9A—O10A—C11A—C12A	-93.7 (2)	C9B—O10B—C11B—C12B	-91.7 (2)
O10A—C11A—C12A—C17A	44.1 (3)	O10B—C11B—C12B—C17B	39.8 (3)
C11A—C12A—C17A—C18A	-1.2 (4)	C11B—C12B—C17B—C18B	-2.6 (4)
C9A—N1A—C18A—C17A	-77.2 (3)	C9B—N1B—C18B—C17B	-81.5 (3)
C12A—C17A—C18A—N1A	23.5 (4)	C12B—C17B—C18B—N1B	30.5 (4)

Although most of the H atoms were observed in a difference Fourier map, all were refined with fixed geometry, riding on their carrier atoms, with U_{iso} set to $1.2U_{\text{eq}}$ of the parent atom.

Data collection: *XSCANS* (Siemens, 1991); cell refinement: *XSCANS*; data reduction: *XSCANS*; program(s) used to solve structure: *SHELXS97* (Sheldrick, 1997); program(s) used to refine structure: *SHELXL97* (Sheldrick, 1997); molecular graphics: *ORTEPII* (Johnson, 1976); software used to prepare material for publication: *SHELXL97*.

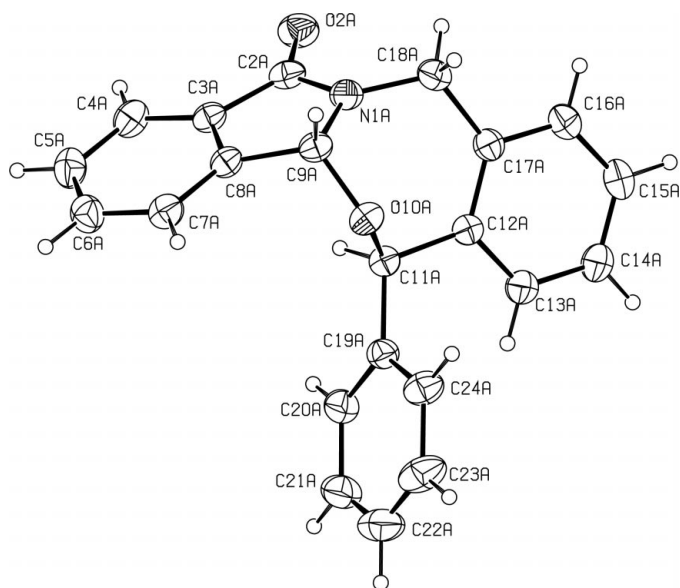


Figure 1
 A view of the title molecule, showing the labelling of the non-H atoms. Displacement ellipsoids are shown at the 30% probability level. For clarity, only molecule A is shown.

This work was supported by the Grant Agency of the Slovak Republic (project Nos. 1/8216/01 and 1/6095/99).

References

Barrett, D. M. Y., Kahwu, I. A., Mague, J. T. & McPherson, G. L. (1995). *J. Org. Chem.* **60**, 4946–4953.
 Benedetti, E., Bavoso, A., DeBlasio, B., Pavone, V. & Pedone, C. (1983). *Biopolymers*, **22**, 305–317.
 Cremer, D. & Pople, J. A. (1975). *J. Am. Chem. Soc.* **97**, 1354–1359.
 Huang, Q., He, X., Ma, C., Liu, R., Yu, S., Dayer, C. A., Wenger, G. R., McKernan, R. & Cook, J. M. (2000). *J. Med. Chem.* **43**, 71–95.
 Johnson, C. K. (1976). *ORTEPII*. Report ORNL-3794. Oak Ridge National Laboratory, Tennessee, USA.
 Khan, M. W., Guha, S., Mukherjee, A. K. & Kundu, N. G. (1998). *Acta Cryst.* **C54**, 119–121.
 Lokaj, J., Kettmann, V. & Marchalin, S. (2001). *Acta Cryst.* **C57**, 735–736.
 McNab, H., Parsons, S. & Shannon, D. A. (1997). *Acta Cryst.* **C53**, 1098–1099.
 Nardelli, M. (1983). *Acta Cryst.* **C39**, 1141–1142.
 Sheldrick, G. M. (1997). *SHELXL97* and *SHELXS97*. University of Göttingen, Germany.
 Siemens (1991). *XSCANS*. Siemens Analytical X-ray Instruments Inc., Madison, Wisconsin, USA.